

Rapid Response to Low-Dose Rituximab Following Development of Severe Hemophagocytic Lymphohistiocytosis Due to Epstein-Barr Virus Infection

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Abstract

Viral infection-associated hemophagocytic lymphohistiocytosis is often observed in children. When Epstein-Barr virus is the pathogen, patients can develop serious coagulopathy and pancytopenia. Usually such patients respond to standard steroid therapy, but alternative therapy for steroid-refractory cases is limited. Here we present the clinical observation of a Japanese girl who was successfully treated with rituximab after she failed to respond to standard steroid therapy.

Keywords: Epstein-Barr virus; Hemophagocytic lymphohistiocytosis; Rituximab

Introduction

Primary hemophagocytic lymphohistiocytosis (HLH) is caused by genetic mutations, while secondary HLH can be caused by infection, malignancy or drugs. Epstein-Barr virus (EBV) is the leading pathogen in secondary HLH. EBV usually infects B-cells but not T-cells, resulting in infectious mononucleosis (IM). Activated T-cells release type-1 cytokines to remove EBV-infected B-cells, following which patients may develop fatal HLH. Although most patients respond to steroid therapy, the use of further steroids, cyclosporine and etoposide have been recommended in refractory patients to suppress the serious cytokine release syndrome [1]. However, etoposide is potentially leukemogenic [2, 3]. Rituximab, an anti-CD20 antibody, removes CD20+ B-cells efficiently and has been used

for CD20+ lymphoma and EBV-associated post-transplant lymphoproliferative disorder [4, 5]. Here we report a HLH case successfully treated with rituximab instead of etoposide.

Our case study will add to the current literature providing evidence of an alternative therapy for steroid-refractory HLH, since the efficacy of rituximab has not yet been fully determined.

Case Report

A 15-year-old girl, who was previously well without remarkable past history was referred to our hospital with 5-day long fever. She did not respond to antimicrobial agents before admission. Her parents are Japanese and have enjoyed a middle-class life style; and the family has not been exposed to a harmful environment, nor does it have any inherited diseases. The physical and neurological examinations that were performed on admission were as follows. The patient's body temperature was 39.8 °C, heart rate was 96/min, respiratory rate was 16/min and blood pressure (mm Hg) was 108/58. Pharyngeal erythema, tenderness in the lower abdomen and splenomegaly were found. There are no other abnormal findings. Laboratory data upon admission were as follows: white blood cells (WBC), $1.0 \times 10^9/L$; platelet count, $48 \times 10^9/L$; fibrin and fibrinogen degradation product (FDP), 13.9 mg/L; thrombin and anti-thrombin complex (TAT), 26.8 µg/L; ferritin, 1,921 µg/L and soluble interleukin-2 receptor, 9,090 U/mL (Table 1). Bone marrow was hypoplastic and the monocyte fraction was markedly increased (Fig. 1). These findings indicated severe HLH, and so we started prednisolone (2 mg/kg/day) from day 2 post admission (Fig. 2). We immediately performed a multiplex viral DNA screening test including herpes simplex virus (HSV)-1, HSV-2, varicella-zoster virus, EBV, cytomegalovirus, human herpesvirus (HHV)-6, HHV-7, JC virus, BK virus and parvovirus by polymerase chain reaction. On day 4, she presented systemic edema and respiratory failure (body weight of 47.5 kg at admission increased to 54.5 kg). In addition, leukopenia and coagulopathy progressively worsened; WBC, $0.3 \times 10^9/L$; platelet count, $36 \times 10^9/L$; FDP, 104.6 mg/L and TAT, 80.2 µg/L. Cyclosporine (6 mg/kg/day, per oral) was then added from day 4.

EBV-DNA copy number was extremely high (340,000 copies/mL), and IgM against EBV viral capsid antigen was

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Table 1. Laboratory Data

	Normal range	On admission										Outpatient clinic					
		day 1	day 2	day 4	day 5	day 6	day 7	day 8	day 9	day 11	day 13	day 15	day 18	day 30	day 56	day 126	day 195
WBC (10 ⁹ /L)	3.3 - 8.6	1	1	0.3	1.4	1.9	2.4	3.6	8.4	5.9	4.5	3.1	3.4	3.6	2.9	4.4	3.8
Neutrophil (%)		64	95	94	96	96.5	Nd	95	93	72	68	72	68	74	71	76	69
Lymphocyte (%)		28	4	3	3	3	Nd	2	2	18	13	16	13	20	23	20	26
Monoocyte (%)		7	1	3	1	0.5	Nd	4	5	10	19	12	16	5	4	4	4
Hb (g/L)	116 - 148	137	126	107	104	113	102	97	98	95	99	111	113	124	127	135	132
Reticulocyte (%)	4 - 20	7	9	5	3	3	2	2	3	3	45	103	51	16	10	10	14
Plt (10 ⁹ /L)	158 - 348	48	58	36	33	48	33	47	77	139	235	310	346	178	171	198	195
TB (mg/L)	3 - 12	9.6	7.4	6	10.8	13	8.9	6.6	7.7	7.5	8.7	9.1	10.5	9.9	5.7	6.3	6.3
AST (U/L)	13 - 30	92	143	227	272	247	218	125	84	73	45	40	24	24	22	23	29
ALT (U/L)	10 - 42	70	63	102	154	171	165	126	107	132	106	105	55	21	15	34	38
LDH (U/L)	124 - 222	692	825	1,046	1,297	1,245	901	677	497	420	385	317	238	175	130	156	144
UN (mg/L)	80 - 190	74	79	85	83	103	105	148	134	128	133	210	199	86	122	106	84
Cr (mg/L)	5 - 9	8.1	8.2	6.1	4.7	5.2	5.2	5	5.1	4.6	4.7	5.6	5.3	5.8	5.6	6.4	6
TP (g/L)	65 - 80	63	51	45	43	46	47	51	58	59	67	71	69	73	67	71	67
Alb (g/L)	38 - 53	36	29	24	23	26	27	30	39	39	44	48	48	51	47	49	46
Ferritin (µg/L)	20 - 120	1,921	Nd	Nd	Nd	8,479.9	Nd	3,849.6	Nd	1,842	Nd	307.8	Nd	58	9	6.9	11
sIL2-R (U/mL)	145 - 519	9,090	Nd	Nd	Nd	14,400	Nd	Nd	4,880	Nd	1,680	Nd	434	182	261	248	248
CRP (mg/L)	0 - 2	80.3	66.1	40.1	31.8	18.9	10.3	6.9	4.4	2.5	1.2	<1	<1	<1	<1	<1	<1
ANA	(-)	Negative															
PT ratio	0.9 - 1.1	1.26	1.13	1.18	1.09	0.99	0.94	0.9	0.86	0.9	0.9	0.91	0.93	1	1.02	Nd	Nd
Fibrinogen (g/L)	1.5 - 4.0	3.34	2.77	1.53	1.44	1.22	1.27	1.13	1.26	1.21	1.23	1.46	1.5	286	2.23	Nd	Nd
FDP (mg/L)	<5	13.9	20.8	104.6	67.8	55.3	30.5	18	11.1	7.1	<5	<5	<5	<5	<5	Nd	Nd
D-dimer (mg/L)	<1	13	20	88.5	48.3	45	28.1	15.9	9	5.5	2.7	1.8	<1	<1	<1	Nd	Nd
AT (%)	70 - 130	97	87	82	89	109	120	128	135	144	144	>150	>150	140	132	Nd	Nd
TAT (µg/L)	<3	26.8	59	80.2	52	38.3	21.4	12.4	2.9	2.7	2.9	1.5	1.5	<1	Nd	Nd	Nd
PIC (mg/L)	<1	2.7	3.1	Nd	5.8	6.4	6.3	4.2	3	2	1.4	1.5	<1	<1	Nd	Nd	Nd
Urinalysis																	
Protein	(-)	+/-	1+	1+	1+	1+	+/-	+/-	-	Nd	-	-	-	Nd	Nd	Nd	Nd
Blood	(-)	+/-	3+	2+	1+	-	-	-	-	Nd	-	-	-	Nd	Nd	Nd	Nd
Sugar	(-)	-	-	-	-	-	-	-	-	Nd	-	-	-	Nd	Nd	Nd	Nd
Blood culture		Negative															

Alb: albumin; ALT: alanine aminotransferase; ANA: anti-nuclear antibody; AST: aspartate aminotransferase; AT: antithrombin; Cr: creatinine; CRP: C-reactive protein; FDP: fibrin and fibrinogen degradation product; Hb: hemoglobin; LDH: lactate dehydrogenase; PIC: plasmin-α2-plasmin inhibitor complex; Plt: platelets; PT: prothrombin time; sIL2R: soluble interleukin 2 receptor; TAT: thrombin-antithrombin complex; TB: total bilirubin; TP: total protein; UN: urea nitrogen; WBC: white blood cells; Nd: not determined.

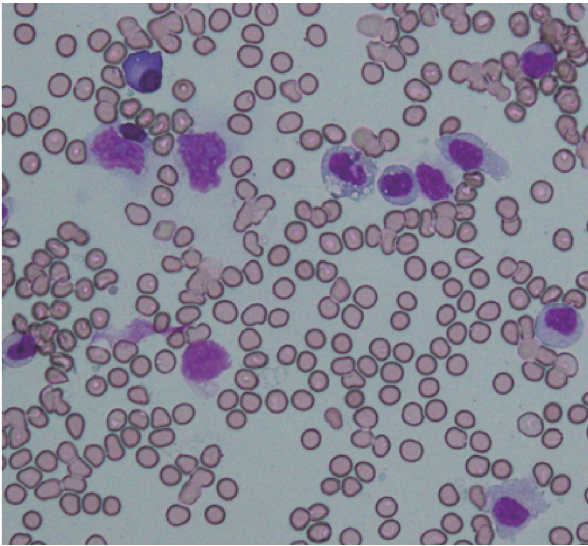


Figure 1. Microscopic findings of bone marrow specimens on day 1. Bone marrow showed monocytic infiltration with a background of hypocellular lesion (May-Giemsa stain, × 400). Lymphocytes account for 12% of total nucleated cells, comprising the following subsets: CD3, 93%; CD4, 36%; CD8, 55%; CD19, 2%; CD56, 4%.

increased but anti-EBV nuclear antigen IgG was not detected. Because these findings indicated a primary EBV infection, we decided to use rituximab to remove EBV-infected cells. On day 5, the first course of rituximab (100 mg/m²) was administered. Surprisingly, her fever and respiratory failure resolved from day 7. WBC and platelet count also began to increase. Coagulopathy and splenomegaly were completely resolved on day 8. Because EBV-DNA copy number still remained at 4,900 copies/mL on

day 9, a second course of rituximab at the same dose was given. On day 16, EBV-DNA was no longer detectable. The patient was discharged at day 20; thereafter prednisolone and cyclosporine were tapered and stopped on day 25 and day 50, respectively. Currently, she is alive and well without any medication.

Discussion

We report a 15-year-old girl who developed severe pancytopenia and coagulopathy due to primary EBV infection and failed to respond to standard steroid therapy. Etoposide has been widely used in the treatment of such refractory cases, but the concern of secondary malignancy development by the use of cytotoxic drug remains. Here, therefore, we employed rituximab, instead of etoposide, to eradicate EBV-infected CD20+ B-cells. Consequently EBV-DNA disappeared completely after two courses of rituximab.

Indeed, it is difficult to discriminate the following disease entities; IM, EBV-HLH, EBV-associated lymphoma and chronic active EBV infection (CAEBV). Specifying the lineage of EBV-infected cells is useful in determining appropriate treatment because EBV usually infects B-lymphocytes in IM, and T or NK-cells in CAEBV. However, the pathogenesis of EBV-HLH is controversial. Although Japanese groups concluded that EBV-infected cells were CD8+ T-cells but not B-cells, other investigators have also reported similar case series as the current patient, which suggests B-cells play a central role [6-10]. In this case, we could not identify the infected cell type; however, we strongly suspect that intrasplenic B-cells were involved in this pathogenesis because all the symptoms and findings such as splenomegaly were quickly resolved upon response to rituximab. Finally, the low-dose rituximab (100

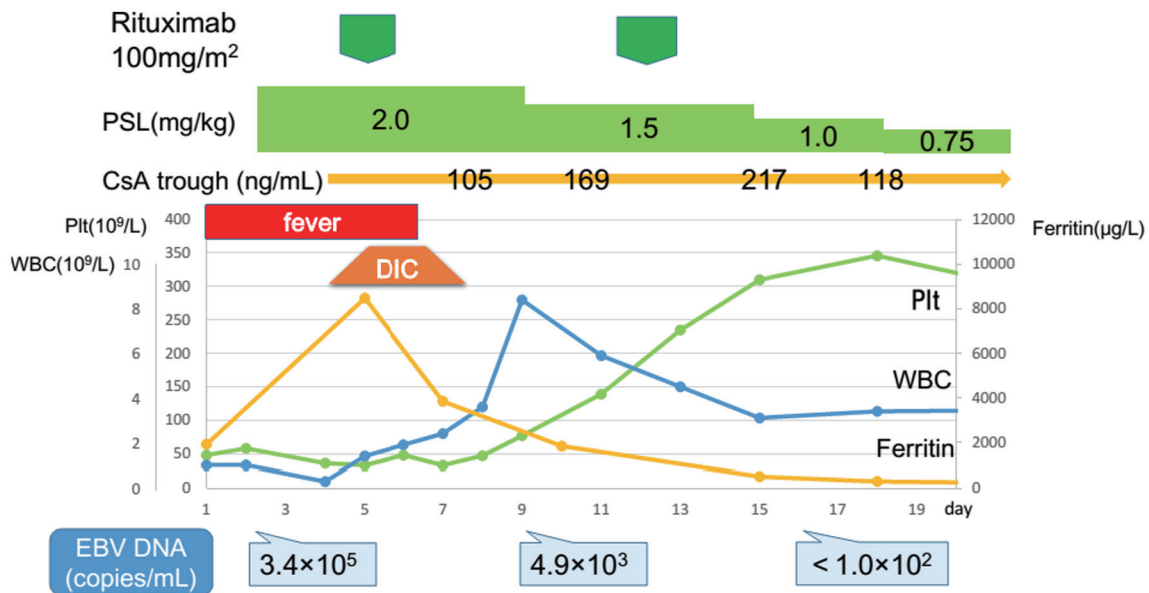


Figure 2. Response to prednisolone, cyclosporine and rituximab. The patient did not respond to first-line therapy with prednisolone. A high level of EBV-DNA was identified on day 4. After the first course of rituximab that was administered on day 5, disease activity rapidly declined. CsA: cyclosporine; DIC: disseminated intravascular coagulation; EBV: Epstein-Barr virus; Plt: platelets; PSL: prednisolone; WBC: white blood cells.

mg/m²) therapy that we used in this patient has been reported in another study for immune thrombocytopenic purpura [11]. Thus the patient could be treated without any adverse effects such as infusion reaction. In summary, eradicating EBV-infected B-cells with two courses of low-dose rituximab successfully cured EBV-HLH in this patient.

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Financial Disclosure

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Conflict of Interest

All the authors declare no conflict of interests.

Informed Consent

All procedures performed in this study were in accordance with the ethical standards of the institution, and the 1964 Helsinki declaration and its amendments. Based on the approval of the institutional review board, written informed consent was obtained from the guardians of the patient for publication of this case report and for any accompanying images.

Author Contributions

HN and HY interpreted the clinical data and performed literature searches. HN prepared the manuscript. HM provided medical care, HY and IM reviewed the manuscript critically and supervised the study. All authors have read and approved the final manuscript.

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